

Stable Transfection of the Bovine *NRAMP1* Gene into Murine RAW264.7 Cells: Effect on *Brucella abortus* Survival

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Genetically based natural resistance to brucellosis in cattle provides for novel strategies to control zoonotic diseases. Bovine *NRAMP1*, the homologue of a murine gene (*Bcg*), has been identified as a major candidate for controlling the in vivo resistant phenotype. We developed an in vitro model for expression of resistance- and susceptibility-associated alleles of bovine *NRAMP1* as stable transgenes under the regulatory control of the bovine *NRAMP1* promoter in the murine RAW264.7 macrophage cell line (*Bcg*⁺) to analyze the regulation of the *NRAMP1* gene and its role in macrophage function. We demonstrated that the 5'-flanking region of bovine *NRAMP1*, despite the lack of TATA and CAAT boxes, has a functional promoter capable of driving the expression of a transgene in murine macrophages. A polymorphism within a microsatellite in the 3' untranslated region critically affects the expression of bovine *NRAMP1* and the control of in vitro replication of *Brucella abortus* but not *Salmonella enterica* serovar Dublin. We did not observe any differences in the production of NO by resting or gamma interferon (IFN- γ)- and IFN- γ -lipopolysaccharide (LPS)-treated transfected cell lines, yet the resistant transfected cell lines produced significantly less NO than other cell lines, following stimulation with LPS at 24 and 48 h.

Losses attributable to infectious diseases continue to impede the livestock industries, despite widespread application of antimicrobials, vaccination, quarantine, and other traditional disease control measures. An alternative approach to the control of zoonotic infections in cattle is genetic disease resistance, which is the inherent capacity of a previously unexposed animal to resist a virulent challenge (1,34). Genetic studies with mice have demonstrated that a gene called natural resistance-associated macrophage protein gene 1 (*Nramp1*, formerly the *Lsh/Ity/Bcg* gene) controls innate resistance and susceptibility to *Mycobacterium bovis* BCG, *Leishmania donovani*, *Salmonella enterica* serovar Typhimurium, and several atypical mycobacteria (D. J. Bradley, Letter, *Nature* **250**:353–354, 1974; 23, 27, 48). The *Nramp1* gene mediates activity of macrophages against intracellular parasites during the early stages of infection (6).

In cattle, natural resistance to *Brucella abortus* is heritable, and its frequency (20% of cross-bred cattle) can be increased dramatically in one generation of selective breeding; the genetic analysis of classical breeding studies is consistent with two or more genes controlling the resistant phenotype (58). Extensive similarities between bovine macrophage function and resistance to brucellosis and murine macrophage function and resistance to *Salmonella*, *Mycobacterium*, and *Leishmania* have been documented (9, 33, 49, 57). Specifically, macrophages from cattle that are genetically resistant to a stringent in vivo challenge with virulent *B. abortus* S2308 are superior in their capacity to control the intracellular replication of not only *B.*

abortus but also *Salmonella enterica* serovar Dublin and *M. bovis* BCG (52). The bovine homologue to the murine *Nramp1* gene (designated bovine *NRAMP1*) was cloned, and the cDNA was sequenced (21). It is predicted to encode a protein that has 12 transmembrane segments with one hydrophilic N-terminal region containing an SH3-binding motif located at the cytoplasmic surface. The gene product contains one potential N-linked glycosylation site and four protein kinase C phosphorylation sites on serine. A 20-amino-acid transport motif is located between the predicted transmembrane domains 8 and 9 and is conserved in murine and human *Nramp1*. Comparison of human and murine *Nramp1* and bovine *NRAMP1* predicted protein sequences indicates a remarkable degree of homology (86.9% identical residues between murine and bovine; 88.6% identical residues between human and bovine). Bovine *NRAMP1* is expressed primarily in macrophages of the reticuloendothelial system. The point mutation within the coding region of murine *Nramp1* which distinguishes the resistant and susceptible alleles of the mouse *Bcg* gene (59) has never been documented in cattle or other mammalian species in which the *Nramp1* homologue has been sequenced. However, in cattle, single-stranded conformational analysis (ssCA) disclosed a highly significant ($P = 0.0089$) association of a polymorphic (GT)_{*n*} microsatellite (where *n* is 13, 14, 15, or 16) located in the 3' untranslated region (3'UTR) of bovine *NRAMP1* with cattle naturally resistant to brucellosis (1). Of the original 11 in vivo-resistant cattle tested by ssCA, 9 were homozygous for the (GT)₁₃ allele while 2 had the (GT)₁₃-(GT)₁₄ genotype. By contrast, of the original 11 in vivo-susceptible cattle tested, only 2 were homozygous for the (GT)₁₃ allele while the others had at least one (GT)₁₄, (GT)₁₅, or (GT)₁₆ allele. These data support the hypothesis that bovine *NRAMP1* is one of the

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major genes determining resistance and susceptibility to brucellosis.

Considering the multigenic determination of resistance and/or susceptibility to bovine brucellosis, we designed an experiment that would isolate the bovine *NRAMP1* gene from the overall genetic background of cattle. The purpose of this study was to transfect the resistance-associated allele and one of the susceptibility-associated alleles of bovine *NRAMP1* into the murine RAW264.7 (*Bcg^s*) macrophage cell line, their expression being driven by the bovine *NRAMP1* promoter. Stable transfectants were analyzed for their expression of the bovine transgene and their capacity to control the intracellular replication of *Salmonella* serovar Dublin and *B. abortus*, prototype intracellular pathogens, as well as their ability to produce nitric oxide (NO) following stimulation with lipopolysaccharide (LPS) and gamma interferon (IFN- γ).

MATERIALS AND METHODS

Parental cell line. RAW264.7 cells, a mouse macrophage (*Bcg^s*) cell line established from the ascites of a tumor induced in a male BALB/c mouse by the intraperitoneal injection of Abelson murine leukemia virus, were purchased from the American Type Culture Collection (Rockville, Md.). The cells were grown in RPMI 1640 (Life Technologies, Gaithersburg, Md.) supplemented with 10% fetal calf serum (HyClone Laboratories, Logan, Utah) in 25-cm² culture flasks (Corning Costar Corporation, Cambridge, Mass.).

Transformation protocol. Twenty-four hours prior to electroporation (56), the cells were split 1:6 in fresh culture medium. Five million cells were added to 10 μ g of linearized plasmid DNA in a total volume of 50 μ l of phosphate-buffered saline and transferred to a 0.4-cm gap electroporation cuvette (BTX, San Diego, Calif.). The cuvettes were left to equilibrate at room temperature for 5 min before electroporation at 750 V/cm, 1,050 μ F, and 129 Ω in an ECM 600 electroporation system (BTX). Dilutions of the transfected cells were seeded in 90-mm petri dishes and allowed to recover for 48 h when selection was applied with G418 (Life Technologies) at a concentration of 400 μ g/ml. Approximately 10-day-old colonies were isolated with cloning rings (Bellco Glass, Inc., Vineland, N.J.) and expanded.

Isolation of the bovine *NRAMP1* promoter. A bovine bacterial artificial chromosome (BAC) library prepared from a purebred Angus bull in the pBeloBac 11 vector (8) was kindly provided by Scott Davis, Department of Animal Science, Texas A&M University. It was screened using a PCR-based strategy with a primer pair designed from the third exon of bovine *NRAMP1*. Positive clones were digested with *NotI* and run on a 1% agarose gel to document the size of the insert. Then, a Southern blot prepared with DNA from positive clones digested with *EcoRI*, *HindIII*, and *EcoRV* was hybridized with a probe homologous to the third exon of bovine *NRAMP1*. The same blot was also hybridized with a 1.2-kb probe, which is composed of the second exon and intron and the third exon, to identify a digestion fragment containing the 5'-flanking region of bovine *NRAMP1*. A 2.6-kb *HindIII* fragment was detected and subcloned into pBlue-script, and both strands of the DNA were sequenced by the dideoxy method (53).

Construction of p Δ 3Bo-GFP. All restriction enzymes were purchased from Promega (Madison, Wis.) or Boehringer Mannheim (Indianapolis, Ind.). All plasmids were transfected into chemically competent *E. coli* (Stratagene, La Jolla, Calif.), and colonies were screened by blue-white phenotyping and/or restriction digestion; mini- and maxipreparations of plasmid DNA were performed using commercially available kits (Promega; Qiagen, Chatsworth, Calif.). Gel purifications of DNA were performed using the QIAEX II Gel Extraction Kit (Qiagen).

An expression vector carrying the green fluorescent protein (GFP) reporter gene under the control of the bovine *NRAMP1* promoter (p Δ 3Bo-GFP) was constructed in pcDNA3 (Invitrogen, San Diego, Calif.). A 734-bp cassette containing the humanized GFP locus was excised from pGreen Lantern (Life Technologies). The 2,138-bp fragment of the bovine *NRAMP1* promoter region used to construct all expression vectors in this study contained 1,484 bp 5' upstream of the transcription start site as well as the first intron. The construct encoding a fusion protein between the 5' end of bovine *NRAMP1* and GFP was cloned into pcDNA3 in which the endogenous viral promoter (pCMV) had been deleted.

Construction of p Δ 3Bo-R. An expression vector carrying the resistance-associated allele of the bovine *NRAMP1* gene under the control of the bovine *NRAMP1* promoter (p Δ 3Bo-R) was constructed in pcDNA3. The cDNA encod-

ing the resistance-associated allele of the bovine *NRAMP1* gene was cloned downstream of the bovine *NRAMP1* promoter fragment; the resulting construct was inserted into pcDNA3 in which the pCMV promoter had been deleted.

Construction of p Δ 3Bo-S16 by site-directed mutagenesis. The (GT)₁₃ microsatellite in the 3'UTR of the resistance-associated allele of the bovine *NRAMP1* gene was mutated into a (GT)₁₆ repeat by site-directed mutagenesis (18). The entire cDNA encoding the resistance-associated allele of *NRAMP1* was subcloned into pT7 Blue (Novagen, Madison, Wis.). A 5'-phosphorylated selection primer was designed from the pT7 Blue sequence so that it would convert the unique *XmnI* site into an *EcoRV* site (Select-P: 5' CCCGAAGAACGATATC CAATGATGAGC 3'). On the same strand of the plasmid template, a 5'-phosphorylated mutagenic primer (Mut-P: 5' CCCTCTCCGTCTTGCTGTGC ATGCACACACACACACAC CACACACACACACACCCTTGCTCTGG CAGGCCAGTG 3') was designed to insert three additional GT repeats in the (GT)₁₃ microsatellite. One hundred ng of plasmid DNA was denatured at 100°C for 3 min in the presence of 100 ng of both the selection and mutagenic primers in a total volume of 20 μ l containing 20 mM Tris-HCl (pH 7.5), 10 mM MgCl₂, and 50 mM NaCl; the tube was then chilled on ice for 5 min and centrifuged briefly to collect condensation. The mutant DNA strand was synthesized with 2 to 4 U of T4 DNA polymerase (Clontech, Palo Alto, Calif.) and 4 to 6 U of T4 DNA ligase (Novagen) at 37°C for 2 h in a total volume of 30 μ l containing a nucleotide mixture at optimum concentrations; the enzymes were inactivated at 75°C for 10 min, and the reaction mixture was cooled to room temperature. The NaCl concentration in the sample was then adjusted in order to get complete digestion with 20 U of *XmnI* at 37°C for 2 h. Following the digestion, an aliquot of the sample was introduced into chemically competent BMH71-18 *mutS* *Escherichia coli*, a DNA mismatch repair-deficient mutant (Clontech). The transformed bacteria were incubated for 1 h at 37°C in 1 ml of Luria broth and then grown overnight in a total volume of 5 ml of Luria broth containing 100 μ g of ampicillin/ml. DNA from the mixed plasmid pool was extracted using the Wizard Plus Minipreps DNA purification system (Promega). One hundred nanograms of pool plasmid DNA was digested with 10 U of *XmnI* for 2 h at 37°C, and after the addition of another 10 U of *XmnI*, the sample was incubated for an additional 1 h at 37°C. For the final transformation, an aliquot of the reaction mixture was introduced into chemically competent *E. coli*. Several colonies were grown overnight for plasmid DNA extraction; the plasmid DNA was digested in two separate reactions with *XmnI* and *EcoRV* and analyzed on an agarose gel. Clones that had lost the unique *XmnI* site and acquired an additional *EcoRV* site were submitted for automated sequencing to verify the mutated sequence. The plasmid insert from several pooled, mutated clones was then substituted for the corresponding fragment in p Δ 3Bo-R to create p Δ 3Bo-S16.

RNAse protection assays. Plasmid templates used for the generation of antisense RNA probes were purified with the Qiagen Plasmid Maxi kit and linearized with the appropriate restriction enzyme. Following two consecutive phenol:chloroform:isoamyl alcohol extractions and an ethanol precipitation, the pellets were washed with 75% ethanol, air dried, and resuspended in RNase-free distilled water. High-specific-activity, radiolabeled antisense RNA probes were produced using the MAXIscript in vitro transcription kit (Ambion, Austin, Tex.) and gel purified. The bovine *NRAMP1*-R and -S RNase protection assay (RPA) probes were designed from the 3'UTR of the gene and had only 47% homology with the corresponding fragment in the murine *Nramp1* gene. Total RNA was extracted from 10⁷ cultured cells using the RNAqueous phenol-free total RNA isolation kit (Ambion), quantitated at A₂₆₀, aliquoted, and stored at -80°C. RPAs were carried out using the RPA II RNase Protection Assay kit (Ambion), using 15 μ g of total RNA and following the recommendations of the manufacturer. The protected fragments were displayed on 5% polyacrylamide-8 M urea gels that were dried and exposed to X-ray film at -80°C with intensifying screens.

In vitro killing assays. In vitro macrophage killing assays were performed essentially as described (50), using virulent strains of *B. abortus* and *Salmonella* serovar Dublin. The day prior to the assay, all cells were washed in G418-free medium, harvested by scraping, counted, pelleted, and resuspended in fresh G418-free medium. Ten thousand cells were then added to each of three wells of two tissue culture 60-well Terasaki HLA plates (Nunc, Inc., Naperville, Ill.). One plate was used for harvesting bacteria at the starting time (*T*₀) while the other was used at the end of the assay (*T*₆ and *T*₁₂, respectively, for *Salmonella* serovar Dublin and *B. abortus*). The plates were centrifuged at 750 \times *g* for 5 min and incubated at 37°C for at least 18 h. Aliquots of each bacterial stock were stored at -80°C and used only once. The day of the assay, serial dilutions of the inoculum were plated to verify the viability of the bacteria and the concentration of the inoculum. The monolayers of macrophages were then challenged with bacteria at different multiplicities of infection (MOIs); the time allowed for phagocytosis was 30 min for *Salmonella* and 2 h for *Brucella*. Extracellular bacteria were then killed with gentamicin at 25 μ g/ml for 1 h at 37°C. After three

washes were carried out, the bacteria were harvested from one of the plates for bacterial counting at the start of the assay. In the other plate, the cells were washed, refed fresh medium, and incubated at 37°C. At the end of the assay, the bacteria were harvested, and serial dilutions were plated in duplicate. The cells were visually inspected at regular time points during the infection. As controls, bacteria were grown in triplicate wells without macrophages to assess their viability; also, each assay included a control to verify the activity of the antibiotic. Bacterial survival was calculated as the ratio of the total number of viable intracellular bacteria at the end of the assay to the total number of viable intracellular bacteria at the start of the assay. Each killing assay included the parental cells as a reference standard.

Measurement of NO production. The cell lines tested for the production of NO included one each of the bovine *NRAMP1*-R and -S16 transgenic lines, a vector-transfected line, and the parental cells. The cells were seeded in 96-well microtiter plates in triplicate (10^5 cells/well) and left untreated or treated with murine recombinant IFN- γ (1 U/ml; R&D Systems, Minneapolis, Minn.), LPS from *E. coli* (5 ng/ml; Sigma, St. Louis, Mo.), or IFN- γ (1 U/ml) plus LPS (5 ng/ml). Each experiment also included control cells that were pretreated with *N*^G-monomethyl-L-arginine (300 μ M; Sigma) and then stimulated with IFN- γ and LPS. After 24 and 48 h, the amount of NO₂⁻ in the culture supernatants was measured using the Greiss reagent; for each cell line, the total concentration of NO₂⁻ was measured from the mean absorbance at 550 nm of triplicate samples. Three independent experiments were performed.

Nucleotide sequence accession number. The complete sequence of bovine *NRAMP1* is available in GenBank under accession number 321467.

RESULTS

We have produced stable transfectants of RAW264.7 cells, a murine macrophage cell line expressing the resistance- and susceptibility-associated alleles of the bovine *NRAMP1* gene under the control of the bovine *NRAMP1* promoter. We have isolated two plasmid clones containing the genomic sequence of bovine *NRAMP1* from a BAC library. The clones both contained 90-kb inserts and displayed identical restriction patterns, indicating that these independent clones are in fact identical. PCR analysis using primer sets from both the 5' and 3' regions of the bovine *NRAMP1* cDNA indicated that the inserts span the entire *NRAMP1* genomic sequence. A 2.6-kb *Hind*III restriction fragment from one of the clones was subcloned and sequenced; this fragment contained 1,484 bp of sequence upstream of the transcription start site (Fig. 1). The analysis of this 1,484-bp fragment indicated that there were no consensus TATA or CAAT boxes, but there are two striking features of a functional promoter including the following: (i) enrichment in G+C dinucleotides with about 66% overall G+C content upstream of the ATG start codon (within 140 bp of nucleotide position -87 to +53) and (ii) the presence of a number of consensus transcription factor binding sites. In addition, a wide variety of potential transcription factor consensus sequences were found located in the first and second intron. Comparison of the sequence upstream of the transcription start site between human and murine *Nramp1* and bovine *NRAMP1* indicated a high degree of homology between positions -80 and -257 of the 5' promoter region, with 58% and 68% nucleotide identity between bovine and murine and bovine and human genes, respectively. Of particular interest is the fact that several well-documented transcription factor consensus motifs are precisely conserved in this segment, including binding sites for interferon response element (IRE), NF- κ B-interleukin-6, PU.1, polyomavirus enhancer A-site 3, the W element, and NF- κ B.

In a preliminary experiment, we tested the capacity of a 2.2-kb fragment of the bovine *NRAMP1* promoter region containing 1,484 bp 5' upstream of the transcription start site as

well as the first intron to drive the expression of a transgene in murine macrophages. For this purpose, we constructed a fusion protein between the 5' end of the bovine *NRAMP1* gene and a humanized clone of GFP, both under the regulatory influence of the bovine *NRAMP1* promoter construct (p Δ 3Bo-GFP). Very weak GFP activity was observed in several stably transfected RAW264.7 clones under UV illumination. Four of them were harvested and expanded, and the presence or absence of expression of GFP mRNA was determined by RPA (Fig. 2). The message for GFP was easily detectable in all four clones with visual GFP activity.

Next, RAW264.7 cells were transfected with eukaryotic expression vectors in which the expression of the resistance- and susceptibility-associated alleles of bovine *NRAMP1* is driven by the same bovine *NRAMP1* promoter region (p Δ 3Bo-R and p Δ 3Bo-S16, respectively). Sixteen stably transfected clones of each region were harvested and expanded. The expression of the transgene was determined by RPA in eight randomly selected clones. When we tested eight *NRAMP1*-R stable transfectants by RPA (Fig. 3), five of the eight clones tested displayed a readily detectable signal for bovine *NRAMP1* after a short exposure; after an overnight exposure, all eight clones were positive, albeit expressing various levels of the transgene. Three clones (R2, R5, and R6) expressing the highest levels of the transgene were selected for further analysis.

The construct encoding the susceptibility-associated allele (p Δ 3Bo-S16) was derived from p Δ 3Bo-R by site-directed mutagenesis. We chose to use the (GT)₁₆ variant of the microsatellite because (i) in a previous analysis of the association between in vivo resistance and susceptibility of outbred and unrelated cattle to experimentally induced brucellosis and the documented polymorphism within the polymorphic microsatellite in the bovine *NRAMP1* 3'UTR, 4 of 11 susceptible cattle had the (GT)₁₆ allele while none of the resistant cattle displayed this allele and (ii) the (GT)₁₆ allele is the longest polymorphic variant of this microsatellite that has been documented in cattle to date. The resulting construct (p Δ 3Bo-S16) was used to transfect RAW264.7 cells, and the expression of the transgene was determined by RPA in eight randomly selected clones; because of the low level of expression of the transgene, the RPA was repeated on those clones expressing the highest levels of *NRAMP1*-S16 (Fig. 4). The level of expression of the bovine *NRAMP1*-S16 gene was three- to sixfold lower than that observed in the *NRAMP1*-R clones, thus suggesting a critical role for the polymorphic microsatellite in determining the level of steady-state mRNA accumulation of *NRAMP1*. The three clones expressing the highest levels of bovine *NRAMP1*-S16 (S5, S6, and S7) were selected for further analysis.

We then used highly standardized in vitro killing assays to assess the capacity of the parental cells and the transgenic lines to control the intracellular replication of a highly virulent strain of *Salmonella* serovar Dublin. The cells were infected with various MOIs (5, 10, and 20), and bacterial survival was determined after 6 h. The results indicate that (i) the parental cells are permissive for *Salmonella* serovar Dublin replication and (ii) the *NRAMP1*-R and *NRAMP1*-S16 clones, while superior to the parental cells in their capacity to control *Salmonella* serovar Dublin, restrict the intracellular growth of *Salmonella* serovar Dublin equally well (Fig. 5). At all MOIs used, we did

AP1 PU.1 IRE-2

-1484 gctttggtcctcaagtttttgaactcaacctgaggaacacacatccagatcaagcctgggttgggaacacgctgattttctct

IRE-1 IRE-2 IRE-2 IBP-1

-1398 gtgtctcaggtccagacttttgagttggaacacccatgacaattcagagtttaaatctcttctggcagaaaagaaaggaacagga

IBP-1

-1312 tagaggacatgagtggtggtgaggaagactctgagggaggttaagtggtgggataaagagttggcagctggcaggagaag

IRE-2

-1226 agattgggagctggaaggtctagagctaacagattgatcatcctaaagtgaatctgcccatagaacgcttctgcttaggtcacc

GM-CSF IRE-1

-1140 caatatttgcaaacatttgagccagttatcaaacagcatatgttgataaaatttccagcatccttgacaaaacagacgactta

IRE-1

-1054 gtgacagtggaccacatcctcacatgacaactgggtggaactctgtgagcactgcctcttcagacagagtttatgacctttgtc

AP2 IRE-1

-968 tccatcatccctccctccctccatagctccatcggttaagaatctgcctgcaatgagagagacctgggtcgattctgggtg

IRE-2 IRE-2 IRE-1 NF-kB IRE-2

-882 ggaagaaacccctggagaaggaatggcaaccactccagttattcttgcctggaaatcccatggacagagagctggcgggtctac

IRE-1 AP1

-796 gtcctatgaggtcgcaagtgttgacacagacttagtgactaaaccaccactccctattacttccccctgatactgaggtgagc

GRE

-710 atcgtttgtcattgagcactgagcagcactgtttcttctcattcccaacctgtctcatagtttatctacctggcctccttg

-624 ggcaccttttgaccttttagctagactgaaccttcttcaacagaaggaacaaggtcagagacaagacgtgatctggtgacaatgtc

IRE-1 AP2

-538 aagtggatcggtggcagaaccaagaccagaattccagtgccctcttcttctgggatctcttggcctgaatcttcagtgatagtc

IRE-1

-452 ccggtggcagtagccggaagatcttccctctaaggcactcagtgggacactgagacctagacagcctcagaactccctccctcc

IBP-1

-366 ccgtggccatgggaccacagattaaagaggaaggaacctgactgcttgtagaggtgaggtgggctagcgggcagtcacgtgat

IRE-1 IBP-1

-280 ctacacatgcatgcccaagtggcaggaagtgttccaaatcagagctaaacttgaggaggtgcagaactcagagtgctcgggaagg

W element GRE PEA3

-194 agccagaggtgttgtgggctcaggccagggggaacaaatgtccttgctgtggccaccagaccccttccctccagaactggccac

NF-IL-6 IRE-2

-108 ttctccttttgaaagtggttttcacaacgtgcctgcgtgtgaggacagctgaggatggctaagagggcgtggaccgcacacatcgcc

+1

PU.1 -> Transcription start site

-22 accgcctagagaggaagcagggGCTTGCCATGCCCGTGAGGGGCTGCCCGGCACGCCAGCCACTCGCACAGAGAG 53

M S

TGCCCCGAGCCTGCGGTCCTCATGTTCAGtgtagcagccccctccaggggtacagagcctgagtggtgggtgtgatggggagagt 134

IBP-1 IRE-1

cagggcggttgaaggaagtcctctgctgagtggtgggtctgcatctcaaccacaatccagacaccagcctggggctgggtcttc 220

IRE-1 PEA3

acacctccaagggcagagccttgctctgtgaacctggaagccttgcaagccttctgctctgctcaggcgccagggaacagcagc 306

tggtggtgctcagtagcctgctcggaacccacgtccctctccacatcctgcgtggagagagaagtagatctgggaatctctgccc 392

tcccatggaaggaggaacctgaaggtccacttctccacgtctactggagtgctggttccatgggtggaggtgatggaccaca 478

IRE-2 GM-CSF IRE-1

gatgggaatgaaggtctcttatttgggtgggcttcatcttctctgtctgcaggagcaggtgggaagagacagatagctttaa 564

IRE-2

ccctgggcccagcagaagagggcaggtcactcacaaggtcttggtgggccccttcacagGTGACACGGGCCCCCAAGCAGG 645

GAGGGACCATATGGCTCCATCTCCAGCCCCACCCAGTCCAGAGCCACAGCAAGCACCTCCCGGAGGGACCT 717

IRE-1 IRE-1

ACCTAAGTGAGAAGATCCCCATTCCGGATACAGAATCGgtgagaacgctggaagccttctgaggtctttcaggaatgca 795

ccggatcatcgggatctcaccagcctggggagggagggaggtctctgaagatcagggcagctcctccgggtctctgaacagctg 881

tgacagacgagtggaactggcctcctctgtctcctcaagtgttagctggggcgcttaaaatttctctaaaaaatatacca 967

IBP-1 IRE2 IBP-1 GAS

agtggcttttttaaatcaatctcagctctcctaggaaggggaaggaacctgggatgagtttgctgtgcagcctcagggatctttat 1053

NF-IL-6 GAS IBP-1

aaattttaaggaaggaaccagacaagggcagggagaggtgaagccggcggtggtctctgggacagtggtgtgttagttgc 1139

tcagttgtgtcggactc*---(second intron: 1030 nt)---tccccctccacagGGTACATTACGCTGAGGAAGC 1807

FIG. 1. Sequence of the 5'-flanking region, exon 1, intron 1, exon 2, intron 2, and part of exon 3 of bovine *NRAMP1*. The transcription start site in exon 1 is shown by the first capital boldface G with +1. The putative binding sequence motifs for transcription factors are underlined with the name of the corresponding factor shown. The exon sequences are indicated by boldface uppercase letters. The complete sequence is available in GenBank under the temporary accession number bankit321467.

not observe a significant difference in the uptake of *Salmonella* between the parental and transfected cell lines.

Next, we assessed the capacity of the parental cells and the transfected cell lines to restrict the growth of *B. abortus* S2308, a fully virulent strain. Since in preliminary experiments with a variety of MOIs and times allowed for phagocytosis the RAW264.7 cells proved to be poorly phagocytic for *Brucella*,

we used a standard MOI of 100 and extended the time allowed for phagocytosis from 30 min to 2 h. The results of the *B. abortus* killing assays are summarized in Fig. 6. These results indicate that (i) the bovine *NRAMP1*-R and *NRAMP1*-S16 transgenic cells are superior in their ability to control the intracellular replication of *B. abortus* when compared to the parental cells and (ii) the difference in survival rates between

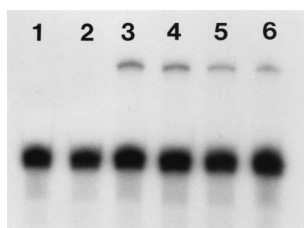


FIG. 2. RPA analysis for expression of GFP under the control of the bovine *NRAMP1* promoter in pΔ3Bo-GFP transfectants of RAW264.7 cells. The message for GFP is easily detectable in all four clones with visual GFP activity (lanes 3 through 6); no signal for GFP is obtained with the parental cells (lanes 1 and 2). The lower dominant band is β-actin. The respective sizes of the protected fragments are 308 (GFP) and 250 (β-actin; positive control) bases. Exposure is 3 h.

the three R and three S16 clones, when analyzed by the Mann-Whitney test, is statistically significant ($P < 0.05$). However, in all the experiments we performed, we also observed marked differences (up to 20-fold) in the number of viable intracellular organisms at T_0 in the parental and transfectant cell lines. While these lower levels of intracellular infection may influence overall bacterial survival in these cell lines, it is important to note that in preliminary assays at similar levels of infection the parental cells consistently had higher rates of survival than those observed in the transfectant lines. In addition, there were reproducible differences in the morphology of the bacterial colonies grown from lysates of the parental cells and the transfectant cell lines at T_0 and T_{12} . While the size of the colonies recovered from the parental cells did not differ from those observed in the bacterial controls or the serial dilutions of the bacterial stock, a majority (60% or more) of the colonies grown from the transgenic cells were reproducibly smaller with a wide distribution of colony sizes ranging from 50 to <10% of expected colony size after 4 days of incubation (Fig. 7). When incubated for an additional 3 to 4 days, the smaller colonies reached normal size. Also, when the smaller colonies were subcultured, the phenotype persisted.

Finally, we assessed the production of NO in the transgenic lines by measuring levels of inorganic nitrite in culture super-

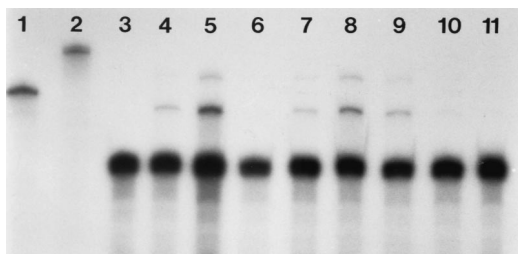


FIG. 3. RPA analysis for expression of bovine *NRAMP1*-R in stable RAW264.7 transfectants. Lane 3, parental RAW264.7 cells; lanes 4 through 11, stable bovine *NRAMP1*-R transfectant clones. A protected fragment of the expected size is observed for the clones in lanes 4, 5, 7, 8, and 9; no signal is obtained with the parental cells (lane 3). The presence of a larger protected fragment can be attributed to the presence of secondary structure in the antisense probe. The lower dominant band is β-actin. The respective sizes of the protected fragments are 289 (*NRAMP1*-R) and 250 (β-actin; positive control) bases. Full-length antisense probes for β-actin and *NRAMP1*-R are shown in lanes 1 and 2. Exposure is 3 h.

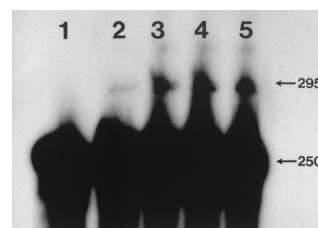


FIG. 4. RPA analysis for expression of bovine *NRAMP1*-S16 in stable RAW264.7 transfectants (lanes 2 through 5). The protected fragments for actin (250 bases) and bovine *NRAMP1*-S16 (295 bases) are indicated. Lane 1, parental cell line. Exposure is 12 h.

natants. In these assays, we compared one each of the bovine *NRAMP1*-R and -S16 transgenic cell lines to vector-transfected cells and the parental cells. The cells were left untreated or stimulated with IFN-γ, LPS, or IFN-γ plus LPS for 24 and 48 h. Only with LPS stimulation at both 24 and 48 h did we observe a difference between the cell lines: the R cell line produced significantly lower levels of NO at both time points than the S16 line and both control cell lines (Fig. 8). We did not observe significant differences between the cell lines under resting conditions or with IFN-γ (data not shown) or IFN-γ-LPS stimulation.

DISCUSSION

Bovine *NRAMP1*, the bovine homologue of a murine gene controlling natural resistance to antigenically and taxonomically unrelated intracellular pathogens, has been identified as one of the major candidates for controlling natural resistance and/or susceptibility to bovine brucellosis in cattle (21). The resistant phenotype in natural resistance to bovine brucellosis is encoded by two or more genes; in addition to bovine *NRAMP1*, *BoLA*, the bovine major histocompatibility complex, is a major candidate gene system believed to control resistance and/or susceptibility to bovine brucellosis. To study the function of bovine *NRAMP1* and because of the multigenic determination of resistance and/or susceptibility to bovine brucellosis, we designed an experimental system in which the bovine *NRAMP1* gene is isolated from the overall genetic background of cattle by transfecting it into the murine RAW264.7 (*Bcg*^s) macrophage cell line, its expression driven by the bovine *NRAMP1* promoter. We selected the RAW264.7 cell line because of its *Bcg*^s background and because it can be easily transfected by electroporation and stable lines can be established (56). In this line of experiments, we cloned and characterized the 5'-flanking region of bovine *NRAMP1*. The 5'-flanking region as well as the first and second introns of bovine *NRAMP1* contain motifs for transcription factors that may play a role in the macrophage-specific expression of *NRAMP1* (NF-κB, NF-interleukin-6, and PU.1) and those related to IFN-γ-inducible gene expression (interferon repressor factor, gamma activation site, and IRE-1 and -2) (20, 22, 37, 42, 44, 60). The high degree of homology between the human and mouse *Nramp1* and bovine *NRAMP1* sequences in the -80 to -257 segment of the 5'-flanking region indicates that this segment may serve as a basal functional promoter (7, 26).

A bovine *NRAMP1*-GFP reporter construct transfected into the RAW264.7 cell line showed only weak GFP activity under

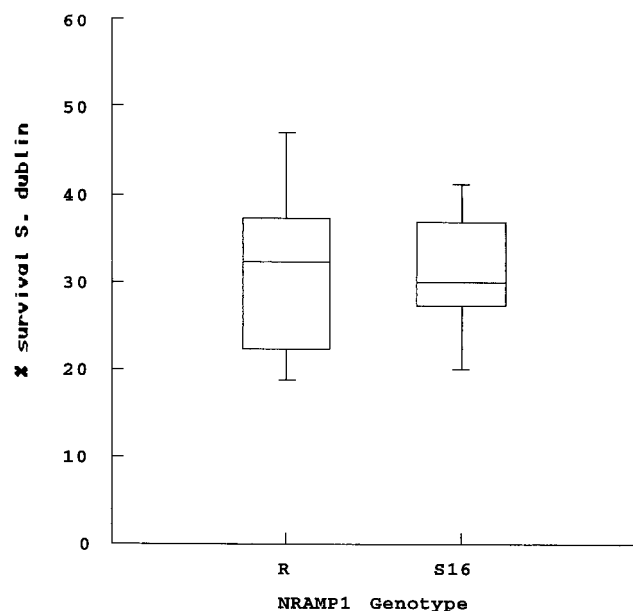


FIG. 5. Survival of *Salmonella* serovar Dublin in three *NRAMP1*-R- or *NRAMP1*-S16-transfected clones of the murine RAW264.7 macrophage cell line. Bacterial survival is expressed as a percentage of survival in the parental RAW264.7 cells. The box plot shows the median and quartiles from two independent experiments; in each experiment, the cell lines were assayed in triplicate, and the survival rates were pooled for the three clones from each genotype. The cells were challenged at a MOI of 20. There is no significant difference in survival rates between the three R and the three S16 clones.

UV illumination, but the mRNA for GFP could be easily detected by RPA. These results indicate that (i) the 5'-flanking region of bovine *NRAMP1* is a functional promoter despite the lack of TATA and CAAT boxes and (ii) the expression of a transgene in the murine RAW264.7 macrophage cell line can be directed from a bovine promoter that has been shown to be active in bovine macrophages. In addition, this promoter construct may be suited for other applications, since it has been shown to be active exclusively in professional phagocytes (12); to this effect, it can be further streamlined by mapping essential elements required for tissue-specific expression. The low level of GFP activity under UV light is attributable to the fact that GFP is a relatively insensitive reporter due to lack of enzymatic amplification (61); about 1 μ M cytosolic GFP (10^5 to 10^6 molecules per cell) is necessary to show up over an autofluorescent background.

Also, it appears that the bovine *NRAMP1* promoter is relatively position independent. This further attribute makes the bovine *NRAMP1* promoter attractive for transgenic applications in which the chromosome position effect may interfere with the expression or inducibility of a transgene (47).

The level of expression of the bovine *NRAMP1*-S16 gene in transfected RAW264.7 macrophages was three- to sixfold lower than that of the resistance-associated allele. Since the *NRAMP1*-R and -S16 constructs were identical except for the presence of three additional GT repeats in the 3'UTR of the S16 construct, it is reasonable to conclude that the polymorphic microsatellite within the 3'UTR of bovine *NRAMP1* may

influence either the rate of transcription or the stability of the bovine *NRAMP1* mRNA. A role for microsatellites in the regulation of gene expression has previously been proposed (D. G. King, Letter, *Science* **263**:595–596, 1994; 17, 32, 43), and the role of the 3'UTR in the posttranscriptional control of mRNA stability and translation has been well documented. Specifically, AU-rich elements have been mapped in the 3'UTR of transcripts encoding cytokines, oncoproteins, and growth and transcription factors (11) and are strongly associated with the regulation of mRNA stability (13,14). In addition, a paradigm for translational regulation by the AU-rich elements has been established for a number of mRNAs (39). More recently, Seiler-Tuyns et al. (54) have provided evidence that, in the case of the human tumor necrosis alpha (TNF- α) mRNA, the 3'UTR also participates in modulating gene expression at the transcriptional level. Because of the random selection process, it is highly unlikely that we selected a whole set of clones that expressed only low levels of the transgene. We cannot exclude the possibility that the promoter region that we used in both constructs may be lacking a regulatory element essential for high levels of expression of the S16 allele of bovine *NRAMP1*; this promoter region was isolated from a BAC clone in which the bovine *NRAMP1* gene had the (GT)₁₃, resistance-associated configuration. Further experiments investigating the rate of transcription of both transgenes as well as the stability of their mRNAs will be necessary to better characterize the role of this polymorphic microsatellite in determining *NRAMP1* gene function. In addition, a thorough

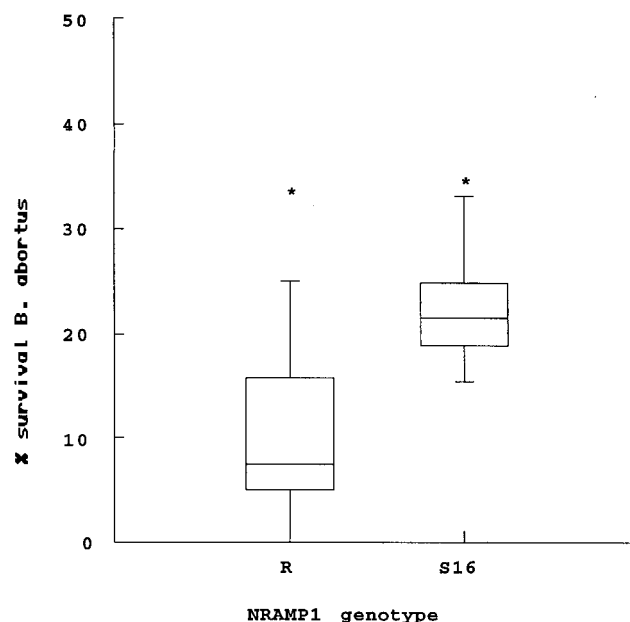


FIG. 6. Survival of *B. abortus* in three *NRAMP1*-R- or *NRAMP1*-S16-transfected clones of the murine RAW264.7 macrophage cell line. Bacterial survival is expressed as the percentage of survival in the parental RAW264.7 cells. The box plot shows the median, quartiles, and outside values (*) from three independent experiments; in each experiment, the cell lines were assayed in triplicate, and the survival rates were pooled for the three clones from each genotype. The difference in survival rates between the three R and the three S16 clones is significant at $P < 0.05$.

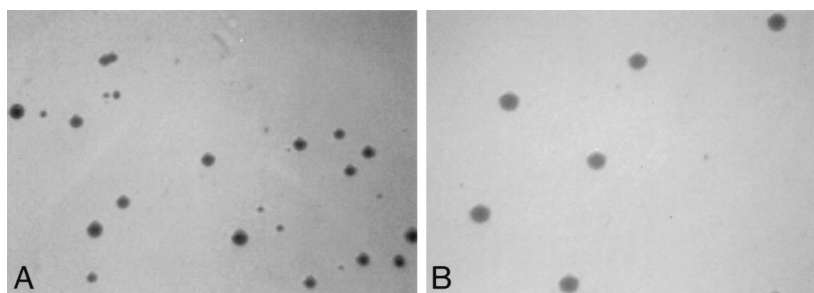


FIG. 7. Colony morphology of *B. abortus* recovered from *NRAMP1*-R transgenic (A) and RAW264.7 (B) cells. *B. abortus* was grown on tryptic soy agar plates for 4 days. Note the normal appearance of *B. abortus* colonies (B) and the wide variation in colony size (A).

analysis of the entire genomic sequence of bovine *NRAMP1* in cattle that are genetically resistant and susceptible to bovine brucellosis is crucial for a better understanding of the regulation of *NRAMP1* expression. We were unable to measure bovine *NRAMP1* protein levels because of the unavailability of an anti-bovine *NRAMP1* antibody of sufficient sensitivity and specificity that would have been suitable for example for a Western blot analysis.

We selected three each of the R and S16 transgenic cell lines for in vitro killing assays, using highly virulent strains of *Salmonella* serovar Dublin and *B. abortus*. In order to minimize interassay variability, the number of clones tested in each assay was limited and the parental, nontransfected cells were included in every assay as an internal control. Thus, we express the survival in each clone tested for each assay as a percentage of the survival obtained with the parental cells in the same assay. When the transfected cell lines were tested for their capacity to control the intracellular replication of *Salmonella*

serovar Dublin, they proved to be significantly superior to the parental cell line at all MOIs. These results indicate that (i) the protein encoded by the bovine *NRAMP1* transgene is functional in RAW264.7 macrophages, (ii) the *NRAMP1* gene plays a role in antimicrobial mechanisms of macrophages directed at controlling *Salmonella* serovar Dublin in vitro, and (iii) the bovine *NRAMP1* gene can complement the mutated murine *Bcg^s* allele.

From these killing assays, it also appears that the polymorphism within the 3'UTR of *NRAMP1*, while strongly influencing the level of steady-state accumulation of the *NRAMP1* mRNA, does not affect the capacity of the S cell lines to control *Salmonella* serovar Dublin in vitro when compared to the R lines. While previous work has demonstrated that macrophages from cattle that are naturally resistant to bovine brucellosis are superior in their capacity to kill both *B. abortus* and *Salmonella* serovar Dublin in vitro when compared to macrophages from susceptible cattle, the existence of a state of

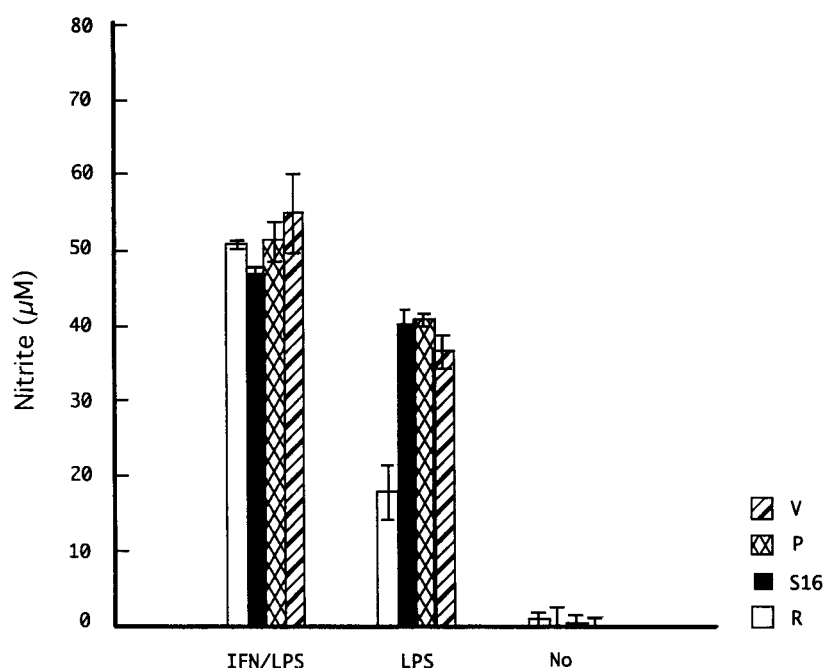


FIG. 8. Nitrite levels in supernatants from bovine *NRAMP1*-R (R), *NRAMP1*-S16 (S16), vector-transfected RAW264.7 cells (V), and parental RAW264.7 cells (P) left untreated (No) or stimulated for 48 h with LPS or IFN- γ -LPS. In each experiment, the cells were assayed in triplicate, and the results are represented as the mean \pm standard deviation of three independent experiments.

genetic resistance to a virulent in vivo challenge with *Salmonella* serovar Dublin has not been documented in these cattle. Our data support the notion that *NRAMP1* plays a role in macrophage killing mechanisms directed at *Salmonella* serovar Dublin but suggest that the difference in in vitro killing observed between cattle that are resistant and susceptible to brucellosis may be unrelated to *NRAMP1*. Considering the importance of *Salmonella* as both a bovine pathogen and a zoonotic agent, future work should be directed at providing a more-thorough understanding of the genetic and molecular mechanisms involved in host defense against *Salmonella* species.

In *B. abortus* killing assays, the microbicidal capacity of the R and S16 cell lines was significantly superior to that of the parental cells, thus suggesting a role for *NRAMP1* in resistance to *Brucella*. Similar to the murine J774A.1 macrophages, adequate levels of infection of the RAW264.7 cells with *B. abortus* were achieved at a high MOI only (35). It has been reported that efficient phagocytosis of *Brucella* by mouse macrophages, contrary to bovine macrophages, requires opsonization by specific antibodies (10, 28). Reproducible differences in the number of viable intracellular organisms at T_0 in the parental and transfected cell lines tend to implicate the bovine *NRAMP1* gene in the process of phagocytosis of *B. abortus*. This finding is not incompatible with the previous demonstration that the differential capacity of macrophages from *Bcg^r* and *Bcg^s* mice to control the intracellular replication of *S. enterica* serovar Typhimurium is linked not to phagocytosis of the organism but rather to increased bacterial killing (41). In our experiments, levels of phagocytosis of *Salmonella* serovar Dublin were comparable in the parental and transgenic cell lines. Also, Radzioch et al. (51), when comparing two widely used bone marrow-derived murine macrophage cell lines, showed that B10S (*Bcg^s*) macrophages phagocytosed *Mycobacterium smegmatis* better at 0.5, 1, and 2 h, but not at 3 or 6 h, than B10R (*Bcg^r*) macrophages. It is very unlikely that the recovery of smaller numbers of viable intracellular bacteria at T_0 from the transfected cell lines is due to the rapid killing of *B. abortus*.

The morphology and growth characteristics of the bacterial colonies grown from lysates of the transgenic cell lines are reminiscent of changes seen in colonies of *B. abortus* when exposed to conditions of acid shock in vitro (40) and may provide important insight into the putative function of the *NRAMP1* protein in that they could implicate *NRAMP1* either directly or indirectly in the phagosome acidification and/or phagosome-lysosome fusion processes inside macrophages. *Nramp1* encodes an integral membrane protein that shares structural characteristics with ion channels and transporters (29). In addition, *Nramp1* has been found not expressed at the plasma membrane but rather localized to the late endocytic compartments of resting macrophages in a Lamp1 (lysosome-associated membrane protein 1)-positive compartment. Upon phagocytosis, *Nramp1* is recruited to the membrane of the phagosome and remains associated with this structure during its maturation to phagolysosome. The targeting of *Nramp1* from endocytic vesicles to the phagosomal membrane supports the hypothesis that *Nramp1* controls the replication of intracellular parasites by altering the intravacuolar environment of the microbe-containing phagosome, possibly by controlling divalent cation concentrations at that site (25). The *Nramp1* gene

has been implicated previously in the process of phagosome-lysosome fusion in murine macrophages when it was shown that the hydrolytic activity of macrophages, as measured by the capacity of lysosomes to fuse with and transfer active hydrolytic enzymes to phagosomes in which *Mycobacterium avium* resides, is an expression of the *Bcg* gene and that this phenomenon is a key antibacterial activity responsible for growth restriction of *M. avium* (15). The generation of an acidic interior is believed to play an essential role in the microbicidal activity of phagosomes (31), and recent work by Hackam et al. (30) suggests that *Nramp1* may directly or indirectly influence the intraphagosomal pH to alter microbial proliferation. Also, Govoni et al. (24) have shown with a transfection model of RAW264.7 macrophages that transfection of wild-type *Nramp1* into *Bcg^s* RAW 264.7 cells abrogates the intracellular replication of *S. enterica* serovar Typhimurium and that the increased antibacterial activity is linked to increased phagosomal acidification.

The survival rate of *B. abortus* was significantly different between the three R and three S16 clones ($P < 0.05$). Thus, it appears that the polymorphic microsatellite affects the rate of transcription or stability of the bovine *NRAMP1* mRNA, which in turn may result in reduced amounts of bovine *NRAMP1*-expressed protein and differential killing of *B. abortus* by cells transfected with the R and S16 alleles of bovine *NRAMP1*. These results are in accordance with the strong association observed between the in vivo phenotype of cattle naturally resistant and/or susceptible to bovine brucellosis and the ssCA polymorphism for bovine *NRAMP1*.

While the putative role of *NRAMP1* in phagosome acidification provides an attractive explanation for our observations, we cannot exclude a possible role for reactive oxygen and nitrogen intermediates, considering their importance in macrophage antimicrobial strategies directed at *Brucella* and *Salmonella* species (2, 16) and the crucial role played by *Nramp1* in the generation of these radicals (6). While *Brucella* is highly susceptible to reactive oxygen intermediates but less susceptible to reactive nitrogen intermediates, the ability of the respiratory burst and nitric oxide synthase to synergistically kill microbial pathogens such as *Salmonella* is well documented. Therefore, we wanted to assess the production of NO in our transgenic cell lines under various stimulation regimens. Surprisingly, both the R and S16 cell lines did not produce higher levels of NO than the two control (*Bcg^s*) cell lines, following any of the treatments. For example, increased nitrite release after IFN- γ -LPS stimulation has been documented in RAW264.7 cells after in vitro transfection of the *Bcg^r* allele (4). However, Radzioch et al. (52) showed with a *Bcg^r* macrophage cell line in which the expression of *Nramp1* had been down-regulated by a ribozyme that the production of NO was markedly lower after IFN- γ stimulation but not if the treatment with IFN- γ occurred in the presence of LPS, suggesting that IFN- γ -but not LPS-induced activation is affected by the inhibition of the *Nramp1* gene. After 24 and 48 h of LPS stimulation, the R cell line produced significantly lower levels of NO than the S16 cell line and the two controls. These results may seem unexpected in light of the superior anti-*Brucella* activity of the *NRAMP1*-R transgenic cell lines. Yet, a series of experiments with a mouse macrophage model of *Brucella suis* infection (28) supports the possibility that in mice, NO favors the elimination

of *Brucella*, provided that IFN- γ and anti-*Brucella* antibodies are present, i.e., following expression of acquired immunity. Furthermore, Jiang et al. (36) showed that while the murine J774A.1 macrophage cell line has anti-*Brucella* activities which are enhanced by IFN- γ activation, it does not produce NO when activated with IFN- γ and infected with *B. abortus*. They also demonstrated that the IFN- γ -induced enhancement of anti-*Brucella* activities by murine peritoneal macrophages was inhibited by the addition of anti-TNF- α antibodies to the cultures, indicating that TNF- α is necessary for full expression of macrophage anti-*Brucella* activities. This is interesting considering that in RAW264.7 cells, endogenous NO has a negative feedback effect on TNF- α synthesis in vitro, mainly by reducing the half-life of TNF- α mRNA (19, 55). These findings underscore the complexity of the relationship between the host and intracellular parasites that cannot be reduced to the expression of a single gene or the production of a single antimicrobial factor. Macrophages utilize a wide array of molecules and mediators in an attempt to control the replication of the parasite, while the pathogen has evolved strategies to evade these antimicrobial mechanisms and avoid detection by the immune system.

Although the bovine *NRAMP1* gene is a major candidate gene controlling natural resistance to brucellosis, it has been shown recently that under field conditions it does not determine resistance and susceptibility to infection with *M. bovis* (3). This finding is in accordance with recent studies with mice that have failed to establish a role for *Nramp1* in protection against virulent *Mycobacterium tuberculosis*, despite the fact that a role for *Nramp1* in controlling multiplication of *M. bovis* BCG vaccine within murine macrophages is well documented both in vivo and in vitro (45, 46). However, a new locus with a major effect on tuberculosis susceptibility, designated *ssl1* (for susceptibility to tuberculosis 1), has been mapped to a 9-centimorgan (cM) interval on mouse chromosome 1, 10 to 19 cM distal to *Nramp1* (38). Similarly, in humans, genetic variants of *Nramp1* account for only a small proportion of the overall genetic component of natural resistance to tuberculosis suggested by twin studies (5).

Further verification of our findings and attempts to repeat them in bovine macrophage cell lines are essential, since they could provide the basis for novel strategies aimed at controlling zoonotic diseases in cattle. Also, our in vitro model should greatly facilitate future work on the regulation and biology of bovine *NRAMP1*.

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REFERENCES

- Adams, L. G., and J. W. Templeton. 1998. Genetic resistance to bacterial diseases of animals. *Rev. Sci. Tech.* 17:200-219.
- Baldwin, C. L., X. Jiang, and D. M. Fernandes. 1993. Macrophage control of *Brucella abortus*: influence of cytokines and iron. *Trends Microbiol.* 1:99-104.
- Barthel, R., J. A. Piedrahita, D. N. McMurray, J. Payeur, D. Baca, F. Suarez Guemes, V. S. Perumaalla, T. A. Ficht, J. W. Templeton, and L. G. Adams. 2000. Pathologic findings and association of *Mycobacterium bovis* infection with the bovine *NRAMP1* gene in cattle from herds with naturally occurring tuberculosis. *Am. J. Vet. Res.* 61:1140-1144.
- Barton, C. H., S. H. Whitehead, and J. M. Blackwell. 1995. Nramp transfection transfers Ity/Lsh/Bcg-related pleiotropic effects on macrophage activation: influence on oxidative burst and nitric oxide pathways. *Mol. Med.* 1:267-279.
- Bellamy, R., N. Beyers, K. P. McAdam, C. Ruwende, R. Gie, P. Samaai, D. Bester, M. Meyer, T. Corrah, M. Collin, D. R. Camidge, D. Wilkinson, E. Hoal-Van Helden, H. C. Whittle, W. Amos, P. van Helden, and A. V. Hill. 2000. Genetic susceptibility to tuberculosis in Africans: a genome-wide scan. *Proc. Natl. Acad. Sci. USA* 97:8005-8009.
- Blackwell, J. M., C. H. Barton, J. K. White, T. I. Roach, M. A. Shaw, S. H. Whitehead, B. A. Mock, S. Searle, H. Williams, and A. M. Baker. 1994. Genetic regulation of leishmanial and mycobacterial infections: the Lsh/Ity/Bcg gene story continues. *Immunol. Lett.* 43:99-107.
- Blackwell, J. M., C. H. Barton, J. K. White, S. Searle, A. M. Baker, H. Williams, and M. A. Shaw. 1995. Genomic organization and sequence of the human NRAMP gene: identification and mapping of a promoter region polymorphism. *Mol. Med.* 1:194-205.
- Cai, L., J. F. Taylor, R. A. Wing, D. S. Gallagher, S. S. Woo, and S. K. Davis. 1995. Construction and characterization of a bovine bacterial artificial chromosome library. *Genomics* 29:413-425.
- Campbell, G. A., and L. G. Adams. 1992. The long-term culture of bovine monocyte-derived macrophages and their use in the study of intracellular proliferation of *Brucella abortus*. *Vet. Immunol. Immunopathol.* 34:291-305.
- Campbell, G. A., L. G. Adams, and B. A. Sowa. 1994. Mechanisms of binding of *Brucella abortus* to mononuclear phagocytes from cows naturally resistant or susceptible to brucellosis. *Vet. Immunol. Immunopathol.* 41:295-306.
- Caput, D., B. Beutler, K. Hartog, R. Thayer, S. Brown-Shimer, and A. Cerami. 1986. Identification of a common nucleotide sequence in the 3'-untranslated region of mRNA molecules specifying inflammatory mediators. *Proc. Natl. Acad. Sci. USA* 83:1670-1674.
- Cellier, M., C. Shustik, W. Dalton, E. Rich, J. Hu, D. Malo, E. Schurr, and P. Gros. 1997. Expression of the human NRAMP1 gene in professional primary phagocytes: studies in blood cells and in HL-60 promyelocytic leukemia. *J. Leukoc. Biol.* 61:96-105.
- Chen, C. Y., F. Del Gatto-Konczak, Z. Wu, and M. Karin. 1998. Stabilization of interleukin-2 mRNA by the c-Jun NH2-terminal kinase pathway. *Science* 280:1945-1949.
- Chen, C. Y., and A. B. Shyu. 1995. AU-rich elements: characterization and importance in mRNA degradation. *Trends Biochem. Sci.* 20:465-470.
- de Chastellier, C., C. Frehel, C. Offredo, and E. Skamene. 1993. Implication of phagosome-lysosome fusion in restriction of *Mycobacterium avium* growth in bone marrow macrophages from genetically resistant mice. *Infect. Immun.* 61:3775-3784.
- De Groote, A. M., U. A. Ochsner, M. U. Shiloh, C. Nathan, J. M. McCord, M. C. Dinauer, S. J. Libby, A. Vazquez-Torres, Y. Xu, and F. C. Fang. 1997. Periplasmic superoxide dismutase protects *Salmonella* from products of phagocyte NADPH-oxidase and nitric oxide synthase. *Proc. Natl. Acad. Sci. USA* 94:13997-14001.
- Deka, R., M. D. Shriver, L. M. Yu, L. Jin, C. E. Aston, R. Chakraborty, and R. E. Ferrell. 1994. Conservation of human chromosome 13 polymorphic microsatellite (CA)_n repeats in chimpanzees. *Genomics* 22:226-230.
- Deng, W. P., and J. A. Nickoloff. 1992. Site-directed mutagenesis of virtually any plasmid by eliminating a unique site. *Anal. Biochem.* 200:81-88.
- Eigler, A., J. Moeller, and S. Endres. 1995. Exogenous and endogenous nitric oxide attenuates tumor necrosis factor synthesis in the murine macrophage cell line RAW 264.7. *J. Immunol.* 154:4048-4054.
- Faist, S., and S. Meyer. 1992. Compilation of vertebrate-encoded transcription factors. *Nucleic Acids Res.* 20:3-26.
- Feng, J., Y. Li, M. Hashad, E. Schurr, P. Gros, L. G. Adams, and J. W. Templeton. 1996. Bovine natural resistance associated macrophage protein 1 (*Nramp1*) gene. *Genome Res.* 6:956-964.
- Fujita, T., S. Ohno, H. Yasumitsu, and T. Taniguchi. 1985. Delimitation and properties of DNA sequences required for the regulated expression of human interferon-beta gene. *Cell* 41:489-496.
- Goto, Y., R. M. Nakamura, H. Takahashi, and T. Tokunaga. 1984. Genetic control of resistance to *Mycobacterium intracellulare* infection in mice. *Infect. Immun.* 46:135-140.
- Govoni, G., F. Canonne-Hergaux, C. G. Pfeifer, S. L. Marcus, S. D. Mills, D. J. Hackam, S. Grinstein, D. Malo, B. B. Finlay, and P. Gros. 1999. Functional expression of *Nramp1* in vitro in the murine macrophage line RAW264.7. *Infect. Immun.* 67:2225-2232.
- Govoni, G., and P. Gros. 1998. Macrophage NRAMP1 and its role in resistance to microbial infections. *Inflamm. Res.* 47:277-284.
- Govoni, G., S. Vidal, M. Cellier, P. Lepage, D. Malo, and P. Gros. 1995. Genomic structure, promoter sequence, and induction of expression of the mouse *Nramp1* gene in macrophages. *Genomics* 27:9-19.
- Gros, P., E. Skamene, and A. Forget. 1981. Genetic control of natural resistance to *Mycobacterium bovis* (BCG) in mice. *J. Immunol.* 127:2417-2421.
- Gross, A., S. Spiesser, A. Terraza, B. Rouot, E. Caron, and J. Dornand. 1998. Expression and bactericidal activity of nitric oxide synthase in *Brucella suis*-infected murine macrophages. *Infect. Immun.* 66:1309-1316.

29. Gruenheid, S., E. Pinner, M. Desjardins, and P. Gros. 1997. Natural resistance to infection with intracellular pathogens: the Nramp1 protein is recruited to the membrane of the phagosome. *J. Exp. Med.* **185**:717–730.
30. Hackam, D. J., O. D. Rotstein, W. Zhang, S. Gruenheid, P. Gros, and S. Grinstein. 1998. Host resistance to intracellular infection: mutation of natural resistance-associated macrophage protein 1 (Nramp1) impairs phagosomal acidification. *J. Exp. Med.* **188**:351–364.
31. Hackam, D. J., O. D. Rotstein, W. J. Zhang, N. Demaurex, M. Woodside, O. Tsai, and S. Grinstein. 1997. Regulation of phagosomal acidification. Differential targeting of Na⁺/H⁺ exchangers, Na⁺/K⁺-ATPases, and vacuolar-type H⁺-ATPases. *J. Biol. Chem.* **272**:29810–29820.
32. Hamada, H., M. G. Petrino, and T. Kakunaga. 1982. A novel repeated element with Z-DNA-forming potential is widely found in evolutionarily diverse eukaryotic genomes. *Proc. Natl. Acad. Sci. USA* **79**:6465–6469.
33. Harmon, B. G., L. G. Adams, J. W. Templeton, and R. Smith. 1989. Macrophage function in mammary glands of *Brucella abortus*-infected cows and cows that resisted infection after inoculation of *Brucella abortus*. *Am. J. Vet. Res.* **50**:459–465.
34. Hutt, F. B. 1958. Resistant big animals, p. 67–99. In *Genetic resistance to disease in domestic animals*. Comstock Publishing Associates, Ithaca, N.Y.
35. Jiang, X., and C. L. Baldwin. 1993. Effects of cytokines on intracellular growth of *Brucella abortus*. *Infect. Immun.* **61**:124–134.
36. Jiang, X., B. Leonard, R. Benson, and C. L. Baldwin. 1993. Macrophage control of *Brucella abortus*: role of reactive oxygen intermediates and nitric oxide. *Cell. Immunol.* **151**:309–319.
37. Klemsz, J. M., S. R. McKercher, A. Celada, C. Van Beveren, and R. A. Maki. 1990. The macrophage and B cell-specific transcription factor PU.1 is related to the ets oncogene. *Cell* **61**:113–124.
38. Kramnik, I., W. F. Dietrich, P. Demant, and B. R. Bloom. 2000. Genetic control of resistance to experimental infection with virulent *Mycobacterium tuberculosis*. *Proc. Natl. Acad. Sci. USA* **97**:8560–8565.
39. Krays, V., O. Marinx, G. Shaw, J. Deschamps, and G. Huez. 1989. Translational blockade imposed by cytokine-derived UA-rich sequences. *Science* **245**:852–855.
40. Lin, J., and T. A. Ficht. 1995. Protein synthesis in *Brucella abortus* induced during macrophage infection. *Infect. Immun.* **63**:1409–1414.
41. Lissner, C. R., R. N. Swanson, and A. D. O'Brien. 1983. Genetic control of the innate resistance of mice to *Salmonella typhimurium*: expression of the Ity gene in peritoneal and splenic macrophages isolated in vitro. *J. Immunol.* **131**:3006–3013.
42. Lowenstein, C. J., E. W. Alley, P. Raval, A. M. Snowman, S. H. Snyder, S. W. Russell, and W. J. Murphy. 1993. Macrophage nitric oxide synthase gene: two upstream regions mediate induction by interferon gamma and lipopolysaccharide. *Proc. Natl. Acad. Sci. USA* **90**:9730–9734.
43. Lue, N. F., A. R. Buchman, and R. D. Kornberg. 1989. Activation of yeast RNA polymerase II transcription by a thymidine-rich upstream element in vitro. *Proc. Natl. Acad. Sci. USA* **86**:486–490.
44. Matsusaka, T., K. Fujikawa, Y. Nishio, N. Mukaida, K. Matsushima, T. Kishimoto, and S. Akira. 1993. Transcription factors NF-IL6 and NF-kappa B synergistically activate transcription of the inflammatory cytokines, interleukin 6 and interleukin 8. *Proc. Natl. Acad. Sci. USA* **90**:10193–10197.
45. Medina, E., and R. J. North. 1996. Evidence inconsistent with a role for the Bcg gene (Nramp1) in resistance of mice to infection with virulent *Mycobacterium tuberculosis*. *J. Exp. Med.* **183**:1045–1051.
46. Medina, E., and R. J. North. 1999. Genetically susceptible mice remain proportionally more susceptible to tuberculosis after vaccination. *Immunology* **96**:16–21.
47. Palmiter, R. D., and R. L. Brinster. 1985. Transgenic mice. *Cell* **41**:343–345.
48. Plant, J., and A. A. Glynn. 1974. Natural resistance to *Salmonella* infection, delayed hypersensitivity and Ir genes in different strains of mice. *Nature* **248**:345–347.
49. Price, R. E., J. W. Templeton, R. Smith III, and L. G. Adams. 1990. Ability of mononuclear phagocytes from cattle naturally resistant or susceptible to brucellosis to control in vitro intracellular survival of *Brucella abortus*. *Infect. Immun.* **58**:879–886.
50. Qureshi, T., J. W. Templeton, and L. G. Adams. 1996. Intracellular survival of *Brucella abortus*, *Mycobacterium bovis* BCG, *Salmonella* serovar Dublin, and *Salmonella typhimurium* in macrophages from cattle genetically resistant to *Brucella abortus*. *Vet. Immunol. Immunopathol.* **50**:55–65.
51. Radzioch, D., T. Hudson, M. Boule, L. Barrera, J. W. Urbance, L. Varesio, and E. Skamene. 1991. Genetic resistance/susceptibility to mycobacteria: phenotypic expression in bone marrow derived macrophage lines. *J. Leukoc. Biol.* **50**:263–272.
52. Radzioch, D., I. Kramnik, and E. Skamene. 1994. Molecular mechanisms of natural resistance to mycobacterial infections. *Circ. Shock* **44**:115–120.
53. Sanger, F., S. Nicklen, and A. R. Coulson. 1977. DNA sequencing with chain-terminating inhibitors. *Proc. Natl. Acad. Sci. USA* **74**:5463–5467.
54. Seiler-Tuyns, A., N. Dufour, and F. Spertini. 1999. Human tumor necrosis factor-alpha gene 3' untranslated region confers inducible toxin responsiveness to homologous promoter in monocytic THP-1 cells. *J. Biol. Chem.* **274**:21714–21718.
55. Sinha, B., A. Eigler, K. H. Baumann, T. F. Greten, J. Moeller, and S. Endres. 1998. Nitric oxide downregulates tumour necrosis factor in mRNA in RAW 264.7 cells. *Res. Immunol.* **149**:139–150.
56. Stacey, K. J., I. L. Ross, and D. A. Hume. 1993. Electroporation and DNA-dependent cell death in murine macrophages. *Immunol. Cell Biol.* **71**:75–85.
57. Templeton, J. W., and L. G. Adams. 1990. Natural resistance to bovine brucellosis, p. 144–150. In L. G. Adams (ed.), *Advances in brucellosis research*. Texas A&M University Press, College Station, Tex.
58. Templeton, J. W., D. M. Estes, R. E. Price, R. Smith III, and L. G. Adams. 1990. Immunogenetics of natural resistance to bovine brucellosis, p. 396–399. In: Allan Teal (ed.), *Proceedings of the 4th World Congress Genetic Applications to Livestock Production*, 23 to 27 July 1990. University of Edinburgh Press, Edinburgh, Scotland.
59. Vidal, S. M., D. Malo, K. Vogan, E. Skamene, and P. Gros. 1993. Natural resistance to infection with intracellular parasites: isolation of a candidate for Bcg. *Cell* **73**:469–485.
60. Xie, Q. W., R. Whisnant, and C. Nathan. 1993. Promoter of the mouse gene encoding calcium-independent nitric oxide synthase confers inducibility by interferon gamma and bacterial lipopolysaccharide. *J. Exp. Med.* **177**:1779–1784.
61. Zlokarnik, G., P. A. Negulescu, T. E. Knapp, L. Mere, N. Burres, L. Feng, M. Whitney, K. Roemer, and R. Y. Tsien. 1998. Quantitation of transcription and clonal selection of single living cells with beta-lactamase as reporter. *Science* **279**:84–89.

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